Mouse anti-Rat IgG1 Secondary Antibody, APC, eBioscience™

Product Details	
Size	100 µg
Species Reactivity	Rat
Host/Isotype	Mouse / IgG1
Class	Monoclonal
Type	Secondary Antibody
Clone	R1-12D10
Conjugate	APC
Excitation/Emission Max	651/660 nm
Form	Liquid
Concentration	0.2 mg/mL
Purification	Affinity chromatography
Storage buffer	PBS, pH 7.2
Contains	0.09% sodium azide
Storage conditions	4° C, store in dark, DO NOT FREEZE!
RRID	AB_2573210

Applications	Tested Dilution	Publications
Flow Cytometry (Flow)	0.25 µg/test	0 Publication
Miscellaneous PubMed (Misc)	-	0 Publication

Product Specific Information

Description: The R1-12D10 monoclonal antibody recognizes rat IgG1 antibodies and can be used as a second step reagent in flow cytometry and microscopy. The monoclonal does not recognize other rat isotypes nor does it crossreact to mouse IgG1.

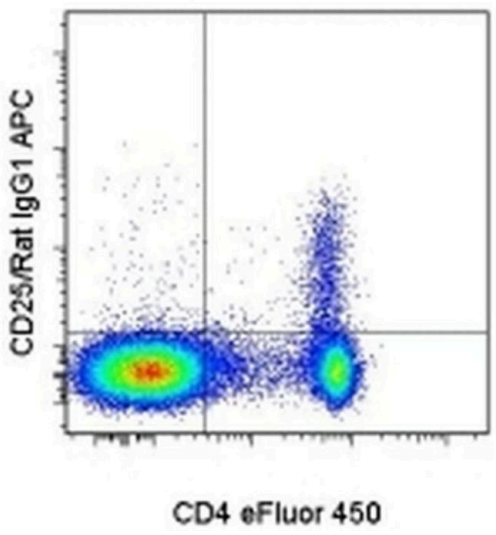
Applications Reported: This R1-12D10 antibody has been reported for use in flow cytometric analysis.

Applications Tested: This R1-12D10 antibody has been tested by flow cytometric analysis of cells stained with a rat IgG1 primary. This can be used at less than or equal to 0.25 µg per test. A test is defined as the amount (µg) of antibody that will stain a cell sample in a final volume of 100 µL. Cell number should be determined empirically but can range from 10^5 to 10^8 cells/test. It is recommended that the antibody be carefully titrated for optimal performance in the assay of interest.

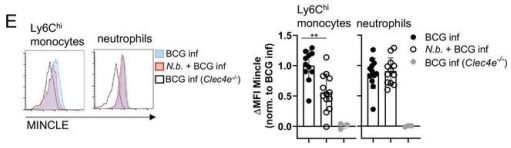
Excitation: 633-647 nm; Emission: 660 nm; Laser: Red Laser.

Filtration: 0.2 µm post-manufacturing filtered.

Product Images For Mouse anti-Rat IgG1 Secondary Antibody, APC, eBioscience™



Rat IgG1 Secondary Antibody (17-4812-82) in Flow
Staining of C57Bl/6 splenocytes with Anti-Mouse CD4 eFluor® 450 (Product # 48-0042-82) and Anti-Mouse CD25 Purified (Product # 14-0251-86), followed by staining with 0.125 µg of Anti-Rat IgG1 APC. Cells in the lymphocyte gate were used for analysis. Quadrant lines were based on the isotype control.



Rat IgG1 Secondary Antibody (17-4812-82) in Flow
Overexpression of IL-4 or co-infection with Nippostrongylusbrasiliensis impair MINCLE upregulation on peritoneal monocytes but does not reduce phagocytosis upon bacille Calmette-Guerin (BCG) infection.(A) IL-4 concentration in serum of mice injected with IL-4 minicircle. C57BL/6 mice were hydrodynamically injected with 0.25 or 0.5 µg of IL-4 plasmid (i.v.) or Ringer solution. Five days (0.5 µg) or 7 days later (0.25 µg) mice were sacrificed and serum IL-4 levels were determined via ELISA (n=5-6 mice per group). n.d.=not detectable. (B, C) 0.25 µg of IL-4 plasmid or Ringer solution was hydrodynamically injected into C57BL/6 wildtype mice. Two days later mice were infected i.p. with 40×106 CFU of Mycobacterium bovis BCG. (B) Generic gating strategy for flow cytometry data. Monocytes were characterized as lineage-CD11b+SiglecF-Ly6C+ cells. Neutrophils were characterized as lineage-CD11b+SiglecF-Ly6G+ cells. Lineage marker: CD3, CD19, NK1.1. (C) Histograms depict MINCLE surface expression on Ly6Chi monocytes and neutrophils 24 hr p.i. analysed via flow cytometry. Quantitative analysis of MINCLE surface expression shown as median fluorescence intensity (MFI). Fluorescence minus one control (FMO) was substracted. Infected MINCLE-/- mice were used as staining controls to exclude unspecific binding of 4A9 antibody. Data is depicted from two independent experiments (2-7 mice per group in total, each dot … Image collected and cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/36753434), licensed under a CC BY license.

6 References

IL-4 and helminth infection downregulate MINCLE-dependent macrophage response to mycobacteria and Th17 adjuvanticity. Elife (2023)

Substratum interactions determine immune response to allogeneic transplants of endothelial cells. Front Immunol (2022)

Regulatory T Cell Depletion Using a CRISPR Fc-Optimized CD25 Antibody. Int J Mol Sci (2022)

Regulatory T Cell Depletion Using A CRISPR Fc-Optimized CD25 Antibody Preprints.org (2022)

Monitoring B cell alloresponses in rats. J Immunol Methods (2022)