Donkey anti-Mouse IgG (H+L) Highly Cross-Adsorbed Secondary Antibody, Alexa Fluor™ 594

Product Details	
Size	1 mg
Species Reactivity	Mouse
Host/Isotype	Donkey / IgG
Class	Polyclonal
Type	Secondary Antibody
Conjugate	Alexa Fluor™ 594
Excitation/Emission Max	590/618 nm
Immunogen	Gamma Immunoglobins Heavy and Light chains
Form	Liquid
Concentration	2 mg/mL
Purification	purified
Storage buffer	PBS, pH 7.5
Contains	5mM sodium azide
Storage conditions	4° C, store in dark
RRID	AB_141633

Applications	Tested Dilution	Publications
Western Blot (WB)	-	0 Publication
Immunohistochemistry (IHC)	1-10 µg/mL	0 Publication
Immunohistochemistry (Paraffin) (IHC (P))	-	0 Publication
Immunohistochemistry (PFA fixed) (IHC (PFA))	-	0 Publication
Immunohistochemistry (Frozen) (IHC (F))	-	0 Publication
Immunohistochemistry - Free Floating (IHC (Free))	-	0 Publication
Immunocytochemistry (ICC/IF)	4 µg/mL	0 Publication
Miscellaneous PubMed (Misc)	-	0 Publication

Product Specific Information

These donkey anti-mouse IgG (H+L) whole secondary antibodies have been affinity-purified and show minimum cross-reactivity to bovine, chicken, goat, guinea pig, hamster, horse, human, rabbit, rat, and sheep serum proteins. Cross-adsorption or pre-adsorption is a purification step to increase specificity of the antibody resulting in higher sensitivity and less background staining. The secondary antibody solution is passed through a column matrix containing immobilized serum proteins from potentially cross-reactive species. Only the nonspecific-binding secondary antibodies are captured in the column, and the highly specific secondaries flow through. The benefits of this extra step are apparent in multiplexing/multicolor-staining experiments (e.g., flow cytometry) where there is potential cross-reactivity with other primary antibodies or in tissue/cell fluorescent staining experiments where there may be the presence of endogenous immunoglobulins.

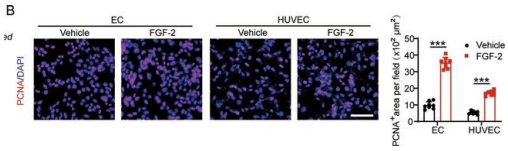
Alexa Fluor dyes are among the most trusted fluorescent dyes available today. Invitrogen™ Alexa Fluor 594 dye is a bright, red-fluorescent dye with excitation ideally suited to the 594 nm laser line. For stable signal generation in imaging and flow

cytometry, Alexa Fluor 594 dye is pH-insensitive over a wide molar range. Probes with high fluorescence quantum yield and high photostability allow detection of low-abundance biological structures with great sensitivity. Alexa Fluor 594 dye molecules can be attached to proteins at high molar ratios without significant self-quenching, enabling brighter conjugates and more sensitive detection. The degree of labeling for each conjugate is typically 2-8 fluorophore molecules per IgG molecule; the exact degree of labeling is indicated on the certificate of analysis for each product lot.

Using conjugate solutions: Centrifuge the protein conjugate solution briefly in a microcentrifuge before use; add only the supernatant to the experiment. This step will help eliminate any protein aggregates that may have formed during storage, thereby reducing nonspecific background staining. Because staining protocols vary with application, the appropriate dilution of antibody should be determined empirically. For the fluorophore-labeled antibodies a final concentration of 1-10 µg/mL should be satisfactory for most immunohistochemistry and flow cytometry applications.

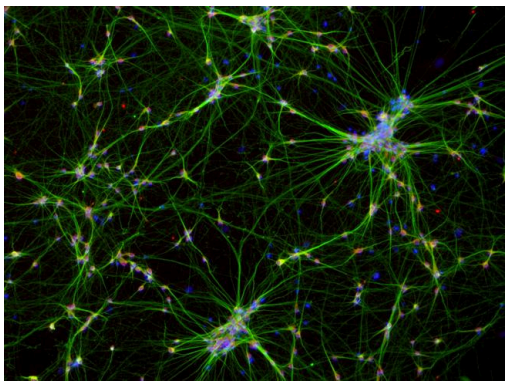
Product will be shipped at Room Temperature.

Product Images For Donkey anti-Mouse IgG (H+L) Highly Cross-Adsorbed Secondary Antibody, Alexa Fluor™ 594



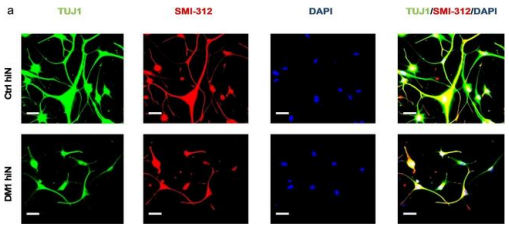
Mouse IgG (H+L) Highly Cross-Adsorbed Secondary Antibody (A-21203) in ICC/IF

FGF-2 impedes the AAD-induced anti-EC effect via FGFR1-ERK-MYC signaling. A Cell growth of human ECs receiving the conditioned medium of scrambled- or FGF2 shRNA-transfected NPC tumor cells (n = 5 samples per group). B Representative micrographs of PCNA+ proliferative cells and DAPI signals in ECs treated with vehicle or recombinant human FGF-2. Scale bar = 50 µm. Quantification of PCNA+ signals in mouse and human ECs (n = 8 random fields per group). C Vehicle- or VEGF-treated ECs were challenged with or without sunitinib or FGF-2. Phosphorylation of AKT and ERK in ECs was detected. -actin marks the loading level in each lane (n = 3 samples per group). D QPCR quantification of Fgfr1, Fgfr2, Fgfr3, and Fgfr4 mRNA levels in ECs (n = 3 samples per group). E Vehicle- or FGF-2-treated ECs were challenged with or without various FGFR inhibitors. Phosphorylation of ERK in ECs was detected. -actin marks the loading level in each lane (n = 3 samples per group). F Downstream of VEGF signaling transcription factors were selected and detected in vehicle- or FGF-2-treated ECs. Heatmap of qPCR array screened out Myc as the highest upregulated transcription factor. G Correlation of FGF2 and MYC transcriptomic expression levels of human NPCs (NPC, n = 113 samples). Data was extracted from dataset GSE102349. H QPCR quantification of Myc mRNA levels in isolated mouse CD31+ ECs from scrambled- or FGF2 shRNA-transfected NPC tu… Image collected and cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/35985991), licensed under a CC BY license.



Mouse IgG (H+L) Highly Cross-Adsorbed Secondary Antibody (A-21203) in ICC/IF

Immunofluorescent analysis of MAP2 in the differentiated neurons from H9 ESC-derived NSCs. 2 weeks after differentiation, cells were fixed, permeabilized and stained with a MAP2 rabbit polyclonal antibody (Product # PA5-17646) at 1:100 dilution (green) and a HuC/HuD mouse monoclonal antibody (Product # A-21271), at a concentration of 5 µg/mL (red) in blocking buffer for at least 1 hour at room temperature, and then incubated with goat anti-rabbit IgG secondary antibody, Alexa Fluor Plus 488 conjugate (Product # A32731, green) and a donkey anti-mouse IgG secondary antibody, Alexa Fluor 594 conjugate (Product # A-21203, red) at a dilution of 1:1000 for 1 hour at room temperature. Nuclei (blue) were stained with Hoechst 33342 dye (Product # 62249).



Mouse IgG (H+L) Highly Cross-Adsorbed Secondary Antibody (A-21203) in ICC/IF

Abnormal axonal outgrowth in DM1 hiNeurons at 15 DPI.(a) Representative fluorescent images display axons of control (top panel) and DM1 (bottom panel) hiNeurons visualized by co-staining with TUJ1 (green) and SMI-312 (red) antibodies. Nuclei stained by DAPI. On the right merged images of TUJ1, SMI-312 and DAPI. Scale bar, 50 µm. (b-e) box and whisker plots show measurements of axonal length of control and DM1 hiNeurons. Results were categorized into four categories: the longest/photo (b), medium length/photo (c), the shortest/photo (d) and average/photo (e). Each sample is shown in different color. Line & (+) sign inside the box represent median and mean of replicates (outcome analyzed), respectively. Whiskers show minimum & maximum values. Note that the average length of axons measured in DM1 hiNeurons group is about half length of axons in control group. n = 3 for each group, a total of 36 (the longest, medium and the shortest/photo analysis) or 108 (average/photo analysis) axons were analyzed per sample. A total of 36 images were analyzed per sample. **P<0.01 compared to control group by unpaired t-test. Average axonal length is the average of the longest, medium and the shortest axons per photo. Medium axon is a single representative of the general population of axons excluding the longest and shortest axons in each photo. Image collected and cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/35776705), licensed under a CC BY license.

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1545 References

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